

PRELIMINARY COMMUNICATIONS

OBSERVATIONS OF 1,1',1''-PHOSPHINOTHIOYLIDYNETRISAZIRIDINE (THIOTEPA)

IN ACIDIC AND SALINE MEDIA. A ¹H-NMR STUDY

Gerald Zon*

Department of Chemistry, The Catholic University of America,
Washington, D.C. 20064

William Egan

Reproduction Research Branch, National Institute of Child Health and Human Development,
National Institutes of Health, Bethesda, Md. 20014

Jerry B. Stokes

Insect Chemosterilants Laboratory, ARS, United States Department of Agriculture,
Beltsville, Md. 20705, U.S.A.

(Received 27 October 1975; accepted 16 January 1976)

Much of the widespread interest in 1,1',1''-phosphinothioylidynetrisaziridine (thiotepa, **1**) derives from its utility as an anticancer agent^{1,2} and chemosterilant.³ Following earlier studies on the acid sensitivity⁴ of thiotepa and possible pharmacological effects⁵ thereof, Benckhuijsen⁶ investigated the behavior of thiotepa under strongly acidic conditions (pH 1.1). It was thus claimed,⁶ on the basis of colorimetric assay data for sulphydryl⁷ and alkylating^{8,9} groups, that protonation of thiotepa results in rapid (< 5 min) rearrangement to **2**, which in turn undergoes relatively slow hydrolysis to **3A** or **3B** (Fig. 1). Thin-layer

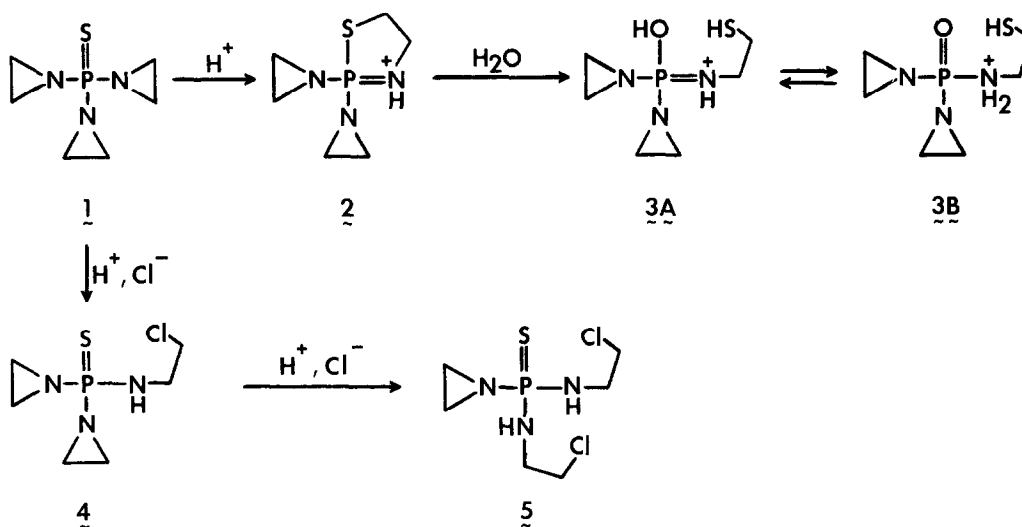


Fig. 1.

* Author to whom correspondence should be addressed.

chromatographic evidence was offered⁶ as proof that aziridine was not formed. In contrast to these conclusions, Maxwell *et al.*¹⁰ have recently suggested that reaction of thiotepa in acidic saline leads to production of 4 and 5, with 4 being the sole product in saline itself. These proposals^{6,10} seemed especially intriguing to us when considered in view of ¹H-NMR results obtained with tepa,¹¹ which clearly demonstrated the vulnerability of phosphorus-bound aziridinyl rings toward acid-catalyzed P-N cleavage and ring-opening. Consequently, we have initiated studies *in vitro* of thiotepa in various media using sensitive high-resolution ¹H-NMR methods to directly elucidate product structures, and now wish to present preliminary findings which disprove the aforementioned claims.^{6,10}

In accordance with the reaction conditions specified by Benckhuijsen,⁶ a freshly prepared solution of purified thiotepa (m.p. 51.5 to 53°; 6.5 mM) in DCl-D₂O (pD 1.2)^{*} was immediately analyzed by ¹H-NMR at 220 MHz (20°) using Fourier transform techniques.¹² The observed spectrum revealed the absence of thiotepa, which would appear in ~ 6 mM D₂O as a doublet (³J_{PH} ≈ 17 Hz) at δ 2.26, as well as the absence of detectable aziridinyl ring proton absorptions attributable to intermediate 2 or final product 3 in the δ 2.0 to 2.5 region.[†] Instead, a sharp singlet at δ 2.67 was accompanied by complex absorption patterns at δ 3.22 to 3.50 and δ 3.62 to 3.91, with the relative integrated intensities of these signals being 1.0 : 2.2 : 1.4. A similar product spectrum obtains for tepa in D₂O buffered at pH 4.0.¹¹ The singlet was identified as N-d₂-aziridinium ion by noting an increase in its relative intensity upon addition of aziridine to the NMR sample. From the observed relative signal intensities, it may be calculated that ~ 20 per cent of the original aziridinyl functionalities in thiotepa were converted to aziridinium ion. While characterization of the remaining reaction products formed under these conditions ([thiotepa] : [2H⁺] = 1 : 10) is still in progress, it has been found (60 MHz) that an analogous product distribution is formed at higher concentrations of thiotepa (~ 60 mM) in the presence of only one molar equivalent of 2H⁺. This observation implies that the aziridinyl rings in thiotepa and products subsequently derived from it are subject to acid-catalyzed P-N cleavage and/or ring-opening. However, with 0.5 molar equivalent of 2H⁺, relative signal integrations showed that ~ 25 per cent of the thiotepa remained; among the usual array of products was a small amount (~ 5-10 per cent) of a new component indicated by a somewhat broadened doublet (J ≈ 17 Hz) at δ 2.16. We are pursuing the isolation and characterization of this latter product, which presumably contains at least one intact aziridinyl ring bonded to phosphorus. The doublet at δ 2.16 was not due

* Correction of a precalibrated standard glass electrode reading of "pH" 0.8 to give an actual value for pD of 1.2 was based on studies by Lumry *et al.*¹³

† All chemical shift values refer to the water-soluble sodium salt of 3-(trimethylsilyl) propionic acid (TSP).

to tepa, since addition of authentic tepa to the reaction mixture gave rise to a relatively low-field doublet ($J \approx 14.5$ Hz) at δ 2.34. It is also noteworthy that sample preparation involving addition of room temperature acid solutions to crystals of thiotepa yields essentially the same spectrum as that obtained by slow mixing of solutions of thiotepa and acid at ice-bath temperatures.

Reaction of thiotepa with one molar equivalent of $2H^+$ in a 4-fold molar excess of sodium chloride gave a product mixture indistinguishable (60 MHz) from that obtained in the absence of added chloride ion. In as much as no high-field absorptions expected for the aziridinyll rings in either **4** or **5** were detected, we are compelled to discard the reaction scheme pertaining to thiotepa in acidic saline proposed by Maxwell *et al.*¹⁰ The additional claim¹⁰ that thiotepa yields **4** in saline was not substantiated by an NMR experiment with 6 mM thiotepa in 1 M sodium chloride: after 2 days no reaction was observed at room temperature; however, at 55° thiotepa did undergo gradual reaction and ultimately afforded (> 3 days) a product spectrum void of an aziridinyll moiety bonded to phosphorus. A control study with thiotepa in pure D₂O at 55° led to similar decomposition, albeit at a somewhat slower rate, thus revealing the competitive incursion of normal hydrolysis. The mechanistic complexity of thiotepa hydrolysis was revealed by ¹H-NMR kinetic measurements in D₂O at 100°, which yielded for the disappearance of thiotepa (0.2 M) an induction period (~ 20 min) followed by adherence to a first-order rate law ($k \approx 3 \times 10^{-4}$ sec⁻¹). By way of comparison, substitution of anhydrous dimethylsulfoxide-*d*₆ (DMSO-*d*₆) for D₂O led to regular first-order decomposition of thiotepa ($k \approx 2 \times 10^{-5}$ sec⁻¹) at 100°, even in the presence of approximately one equivalent of sodium chloride (0.2 M). This thermal instability and resistance to nucleophilic attack of thiotepa in DMSO apparently foiled our attempt to prepare authentic **2** by reaction of thiotepa with sodium iodide¹⁴ in DMSO at 80°, which instead gave insoluble dark colored tarry material. Detailed kinetic and product studies for these reactions of thiotepa will be reported at a later date.

While complete definition of the actual reaction pathways available to thiotepa in acidic media awaits further experimentation, it is none the less apparent that conclusions regarding product structures and mechanisms based on indirect chemical evidence (e.g. functional group tests) must now be considered tenuous. A direct and more reliable investigative methodology may be found in the various NMR spectroscopic techniques.

Acknowledgements. This investigation was supported in part by NIH Grant CA-16158, awarded to G. Z. by the National Cancer Institute, PHS/DHEW. We thank Ms. Maria L. Thomas for manuscript preparation.

REFERENCES

1. J. F. Holland, Cancer Res. 21, 1086 (1961).
2. F. A. Valeriote and S. J. Tolen, Cancer Res. 32, 470 (1972).
3. J. P. Bennett, in Chemical Contraception, pp. 154-7. Columbia University Press, New York (1974).
4. L. B. Mellett and L. A. Woods, Cancer Res. 20, 524 (1960).
5. T. A. Connors, L. A. Elson and C. L. Leese, Biochem. Pharmac. 13, 963 (1964).
6. C. Benckhuijsen, Biochem. Pharmac. 17, 55 (1968).
7. G. L. Ellman, Archs Biochem. Biophys. 82, 70 (1959).
8. R. Truhaut, E. Delacoux, G. Brule and C. Bohuon, Clinica chim. Acta 8, 235 (1963).
9. C. G. Butler, D. S. Kaushik, J. Maxwell and J. G. P. Stell, J. Mond. Pharmac. 4, 359 (1967).
10. J. Maxwell, D. S. Kaushik and C. G. Butler, Biochem. Pharmac. 23, 168 (1974).
11. M. Beroza and A. B. Bořkovec, J. med. chem. 7, 44 (1964).
12. T. C. Farrar and E. D. Becker, in Pulse and Fourier Transform NMR, Introduction to Theory and Methods, p. 1 ff. Academic Press, New York (1971).
13. R. Lumry, E. L. Smith and R. R. Glantz, J. Am. chem. Soc. 73, 4330 (1951).
14. H. W. Heine, W. G. Kenyon and E. M. Johnson, J. Am. chem. Soc. 83, 2570 (1961).